

Study of MiRNA-155 Gene Expression in Egyptian Patients with Chronic Hepatitis C Viral infection

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- Expression
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Abstract

Objective: to evaluate the miRNA -155 expression in patients with chronic HCV infection & correlate it with hepatitis C viral load and treatment response.

Background: MicroRNAs (miRNAs), are small, single-stranded, noncoding RNAs that consist of 20 to 25 base pair (bp). It is a class of small RNAs that regulate mRNA translation and function as oncogenes or tumor suppressor genes. In patients infected with HCV, miRNA-155 expression levels were markedly increased and promote hepatocyte proliferation and tumorigenesis by modulating Wnt signaling.

Subjects and Methods: This study was conducted on 20 HCC patients, 60 chronic liver disease (CLD) patients due to HCV infection subdivided in to 20 patients with HCV naive treatment, 20 patients responder treatment to interferon, 20 patients non responder treatment to interferon patients and 20 healthy subjects matching age and gender. Serum AFP was measured for all participants. The relative expression of miRNA-155 was determined in whole blood samples using Real-time polymerase chain reaction.

Results: The results revealed over expression of miRNA-155 in each of HCC patient group, patients with HCV naive treatment and patients non responder to interferon treatment. However, miRNA-155 showed down expression in patients responder to interferon treatment. miRNA-155 expression was positively correlated with presence of cirrhosis, increased number of focal lesions, larger size of tumor, advanced tumor stage and presence of vascular invasion. From ROC curve analysis, the best cutoff of miRNA-155 chosen to differentiate HCC cases from non HCC subjects was 3.41 RQs (Fold expression), and at this point the sensitivity, specificity, +ve predictive value (PPV), -ve predictive value (NPV) and Accuracy were 88.8%, 91%, 92.4%, 89.5%, 91.4% respectively. For AFP the best cutoff was 85.3 ng/ml at this cutoff point the Sensitivity, Specificity, PPV, NPV, Accuracy were (76.2%, 87.3%, 90.2%, 72.41%, 81.0%). Furthermore, combined use of serum AFP and circulating miRNA-155 for detection of HCC cases, had the advantage over the use of AFP alone as the sensitivity, specificity, PPV, NPV and overall accuracy were increased (89.32%, 91.9%, 93.5%, 90.71% and 92% respectively).

Conclusion: miRNA-155 expression might be correlated with hepatitis C viral load and treatment response. Also, miRNA-155 could be a novel diagnostic and prognostic biomarker for detection of HCC in combination with AFP as well as it could serve as a potential therapeutic target for HCV and HCC infection

INTRODUCTION

HCV was discovered in 1989 as the major causative agent of non-A non-B hepatitis (1) HCV is a major cause of post transfusion and community acquired hepatitis. Approximately 70–80% of HCV patients develop chronic hepatitis of which 20–30% leads to cirrhosis and HCC. Treatment options for chronic HCV infection are limited and a vaccine to prevent HCV infection is not available (2).

Hepatocellular carcinoma (HCC) is the fifth most common cancer worldwide and is associated with liver cirrhosis (LC) in 80% of cases(3).In Egypt, the incidence rate of HCC has increased sharply in the last decade(4).The development and progression of HCC is a complex process, which involves the dysregulation of oncogenes and tumor suppressor genes. It has previously been reported that microRNAs (miRNAs) are essential in oncogenesis by the regulation of oncogenes and tumor suppressor genes (5).MicroRNAs are approximately 22-nucleotide, noncoding, endogenous RNA molecules with an important role in various cellular biological processes, including embryonic development, cell differentiation, and tumorigenesis (6).. In humans miRNA-155 is encoded by the MIR155 host gene or MIR155HG.(7) miRNA-155 plays a role in various physiological and pathological processes (8). Exogenous molecular control in vivo of miRNA-155 expression may inhibit malignant growth (9), viral infections, (10) and enhance the progression of cardiovascular diseases.(11)In patients infected with HCV, miRNA-155 expression levels were markedly increased, and

promote hepatocyte proliferation and tumorigenesis by modulating Wnt signaling (12). Chronic HCV infection induced liver fibrosis is mediated by upregulation of transforming growth factor (TGF)- β (13). recently miRNA-155 was found to be involved in TNF α and monocytes inflammatory activation which suggest a role for miRNA-155 in the immune response to HCV infection (14).

Better understanding of the molecular mechanisms involved in hepatocellular carcinogenesis contributes to identification of novel prognostic and diagnostic biomarkers and therapeutic targets for HCC. So this study aimed to evaluate the miRNA -155 expression in the patients with chronic HCV infection & HCC and correlate it with hepatitis C viral load and treatment response .

1. Subjects and Methods

1.1. Study population

The study was conducted on 100 subjects divided into 3 groups, selected from inpatient wards and outpatient clinic, National Liver Institute, Menoufia University .

The patients were grouped as following:

Group 1 (HCC on top of HCV infection group):

This group included 20 newly diagnosed patients before receiving therapy. The diagnosis was based on clinical examination, laboratory tests, ultrasonography and spiral CT.

Group 2 (Chronic Viral Hepatitis C - HCV Group): This group included

- 20 patients with HCVnaive treatment matching age and gender..
- 20 patients(Responder treatment to Interferon)

- 20 patients (Non Responder treatment to Interferon)

They were diagnosed by ultrasonographical findings (shrunken liver, coarse echopattern, attenuated hepatic vein and fine nodular surface) and biochemical evidence of parenchymal damage as well as liver biopsy in some cases. the dose and duration of antiviral therapy is 180 mcg SC once weekly for 48 weeks

Group 3 (Control group): Included 20 apparently healthy subjects served as a control group,

The criteria for inclusion in this study:

- 1- The diagnosis of HCC was based on triphasic CT or contrast enhanced dynamic MRI.

The presence of typical features of arterial enhancement and rapid portal or delayed washout on one imaging technique was diagnostic of HCC for nodules >2cm in diameter in cirrhotic patients. In cases of uncertainty or atypical radiological finding, diagnosis was confirmed by biopsy .

The criteria for exclusion in this study:

None of the patients had bacterial or other viral infection, chronic renal damage, insulin-dependent diabetes mellitus (IDDM) and other malignant diseases. The patients undergoing immune-suppressive therapy were also excluded from this study.

All patients and control groups were subjected to the following:

1. Complete history taking.
2. Complete clinical examination.
3. Abdominal ultrasonography and or CT.

The study was approved by ethics committee of National Liver Institute, Menoufia University. Enrolment of individuals in the study

was conditioned by an obtained written informed consent.

1.2. Laboratory investigations:

Ten ml venous blood samples were collected from patients and controls and divided into three aliquots; two aliquots used for routine laboratory investigations; liver function tests using fully automated auto analyzer SYNCHRON CX9ALX (Beckman Coulter Inc., CA, USA), CBC using Sysmex K-21, (Sysmex Corporation, Kobe, Japan) and immunoassay; serum HBs-Ag. by using (Abbott Laboratories, Abbott Park, IL, USA) and HCV-RNA by using Real time PCR . Serum AFP concentration was measured using the Automated Chemiluminescence System (ACS: 180 provided by Siemens Medical Solutions Diagnostics Corporation, USA). The third aliquot was collected in EDTA containing tube and used immediately for miRNA extraction and molecular testing.

2.3. Molecular testing:

Quantification of miRNA-155 using Real Time PCR Technology Using 7500 Fast Real Time PCR - TaqMan® microRNA Assay for: MiRNA-155 and its control gene (MiRNA-16)

3. Extraction and cDNA synthesis:

3.1. MiRNA extraction:

MiRNAs were extracted from fresh EDTA treated blood sample using Qiagen miRNA Extraction kit and QIAzol (Lysis solution) according to the manufacturer's instructions. Single-stranded cDNAs were generated using TaqMan® MicroRNA Reverse Transcription Kit .

3.2. Amplification:

Quantitative PCR using the Applied Biosystems 7500 fast real time System (Applied

Biosystems, Foster City, CA) was used to determine levels of miRNA -155(by TaqMan miRNA Assay) using Universal TaqMan master mix according to the manufacturer's protocol. The primers for miRNA-155, miRNA-16, were supplied by Qiagen Germany.miRNA-155Forward Primer

CTCCTTCCTTTCAACAGAAAATGGA

Reverse

PrimerAAAACAAACATGGGCTTGACATTTAA

miRNA 16forward Primer

GCGAATCATTATTTGCTGCTCTAGAAA

Reverse

PrimerGCTCTGTAACAGCTCTGATACTTAACA

3.3.Quantification:

Quantification of gene expression of miRNA-155 was accomplished by measuring the fractional cycle number at which the amount of expression reached a fixed threshold (Ct), which was directly related to the amount of product. The relative quantification given by the Ct values was determined and the control gene Ct subtracted to achieve ΔCt [$\Delta Ct = Ct$ (gene of interest) - Ct (control gene)]. Then relative expression level was determined as $2^{(-\Delta\Delta Ct)}$, where $\Delta\Delta Ct = \Delta Ct$ (target sample) - ΔCt (reference sample).

3.4.Statistical analysis:

Data were collected, tabulated and statistically analyzed by SPSS version 24.0 statistical package (SPSS, Inc, Chicago, IL, USA). ANOVA (followed by LSD post-hoc test), Kruskal Wallis and Pearson's correlation tests were performed at 5% level of significance. The diagnostic performance for miRNA-155 and AFP to discriminate HCC cases from those without HCC was evaluated using Receiver Operating

Characteristic (ROC) curve analysis. The sensitivity, specificity, positive predictive value (PPV) and negative predictive value (NPV) for each marker were determined.

2. Results

A total of 100 subjects were enrolled in this study, 20 HCC patients (86% were males) with mean age of 46.35 ± 5.65 years, 60 CLD patients (75% were males) with mean age of 42.00 ± 6.88 years, and 20 healthy volunteers (90% were males) with mean age of 40.40 ± 6.40 years. The studied groups were homogenous in terms of age and gender ($p > 0.05$).

- **Comparison of miRNA-155 and serumAFP among HCC, HCV (subgroups) and Control groups revealed** that There was significant increase of miRNA-155 in HCC group compared to each of (HCV NAÏVE, HCV responder, HCV Non responder and control group) ($p < 0.01$). As well as there was significant increase of miRNA-155 in each of (HCV NAÏVE and HCV non-responder group compared to control group ($p < 0.01$)). Meanwhile, there was no significant difference between control group and HCV Responder group regarding miRNA-155 ($p > 0.05$). Also, there was highly significant difference between all the studied group regarding AFP ($p < 0.01$). (Table 1 & fig 1)
- **Statistical comparison between clinical and pathological criteria of tumor regarding mean RQ of miRNA-155 in HCC group (No=20).**

The mean RQ of miRNA-155 showed significant increase, with presence of cirrhosis, increased number of focal lesions, larger size of tumor,

advanced tumor stage and presence of vascular invasion(Table 2)

▪ **Pearson Correlation Matrix Between miRNA-155 And different variables in HCC group**

There was significant correlation between miRNA-155 and each of (GGT and AFP)in HCC group. (Table 3 & fig 4,5)

▪ **Pearson Correlation Matrix Between miRNA-155 And different variables in HCV NAIVE group**

▪ There was significant correlation between miRNA-155 and DB in HCV NAIVE group. (Table 4&fig6)

▪ **Pearson Correlation Matrix Between miRNA-155 And different variables in HCV NON Responder group(data not shown).**

There was significant correlation between miRNA-155 and each of (AST and prothrombin concentration) in HCV non responder group.

Receiver Operator of Characteristics (ROC) curve analysis of AFP and/or miRNA-155 in HCC group (No=20) versus non HCC: displayed that the best cut-off of serum AFP for differentiation of HCC cases from those without HCC was 85.3ng/ml at this cutoff point the Sensitivity , Specificity , +ve predictive value , -ve predictive value , Accuracy were (76.2 % , 87.3 % , 90.2 % , 72.41 % , 81.0 %). For miRNA-155 the best cutoff was 3.41RQs and at this point the sensitivity, specificity, +PPV, -NPV and Accuracy were (88.8%91%, 92.4% ,89.5% ,91.4%) Respectively . Furthermore, combined use of serum AFP and circulating miRNA-155for detection of HCC cases, had the advantage over the use of AFP alone as the sensitivity, specificity, PPV, NPP and overall accuracy were increased (89.32 % , 91.9 % , 93.5%, 90.71 % and 92 % respectively).

Table (1):Comparison of MicroRNA -155 and Plasma AFP among HCC, HCV (Groups)and Control groups.

Studied variables	HCC (N=20) Mean ± SD	HCV NAIVE (N=20) Mean ± SD	HCV Responder (N=20) Mean ± SD	HCV Non Responder (N=20) Mean ± SD	Control (N=20) Mean ± SD	Kruska l- Wallis Test	P-Value	Tamhane Post Hoc p-value
Micro RNA 155 RQ (Folds)	15.1± 7.62	2.05± 0. 53	1.11± 0.62	5.75 ± 1.8	1.06 ± 0.18	86.75	< 0.01**	P1= < 0.01** P2= < 0.01** P3= < 0.01** P4= < 0.01** P5 = < 0.01** P6 = > 0.05 P7 = < 0.01**
Plasma AFP (ng/ml)	34.27 ± 22.28	6.27 ± 3.98	11.08 ± 9.34	13.6 ± 9.4	2.5± 0.79	53.4	< 0.01**	P1= < 0.01** P2= < 0.01** P3= < 0.01** P4= < 0.01** P5 = < 0.01** P6 = < 0.01** P7 = < 0.01**

RQ: relative quantity. p<0.01: significant. P1 between HCC and HCV NAIVE. P2 between HCC and HCV Responder. P3 between HCC and HCV NonResponder. P4 between HCC and control. p5 between control and HCV NAIVE. p6 between control and HCV Responder. p7 between control and HCV NonResponder

Table (2): Statistical comparison between clinical and pathological criteria of tumor regarding mean RQ of miRNA-155 in HCC group (No=20).

Studied variables	MiRNA-155 expression (fold change) in HCC group (No = 20) (M ± SD)	Mann-Whitney Test	P value
Cirrhosis			
Positive (No=14)	18.0 ± 7.1	8.0	< 0.01**
Negative (No=6)	8.27 ± 3.11		
Number of focal lesions			
Single (No=7)	8.12 ± 2.86	4.0	< 0.01**
Multiple (No=13)	18.85 ± 6.64		
Size of tumor			
< 3 cm (No=5)	10.3 ± 6.3	16.0	< 0.05*
≥ 3cm (No=15)	16.6 ± 7.5		
Vascular invasion			
Positive (No=12)	19.7 ± 6.14	3.1	< 0.01**
Negative (No=8)	8.18 ± 2.7		
		Kruskal Wallis	
Stage of tumor (TNM)			
1 (No=3)	7.7 ± 4.7	14.28	** < 0.01
2 (No=3)	8.6 ± 1.4		
3 (No=3)	8.0 ± 0.65		
4 (No=11)	20.7 ± 5.2		
Type of tumor			
Trabecular (No=10)	14.3 ± 8.6	1.054	>0.05
Trabecular & acinar (No=8)	15.7 ± 7.5		
Sclerotic (No=2)	17.1 ± 4.2		

p<0.05: significant.
p>0.05: non-significant

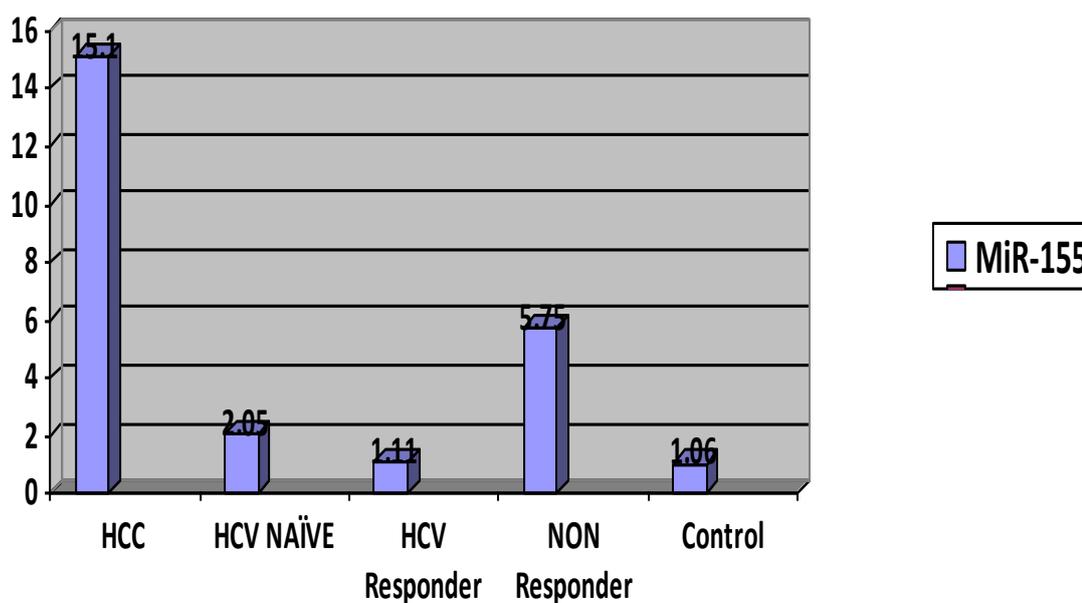


Figure 1: miRNA155 differences between Hepatocellular carcinoma, HCV (NAIVE, responder & non responder) and control groups

Table (3): Pearson Correlation Matrix Between miRNA-155 And different variables in HCC group

Studied variables	HCC (N=20) MiR-155	
	r*	P-Value
AST(IU/L) Up to 40	0.284	>0.05
ALT(IU/L) Up to 42	0.271	>0.05
ALP (IU/L) M Up to115 F Up to 104	0.257	>0.05
GGT (IU/L) 7 – 33	-0.435	<0.05*
T.B (mg/dl) 0 – 1.0	-0.93	> 0.05
D.B (mg/dl) 0 – 0.25	0.98	>0.05
TP (g/dl) 6.5 – 8.5	-0.31	>0.05
Alb (g/dl) 3.5 - 5	-0.205	> 0.05
PC % 80 -120	0.193	> 0.05
AFP (ng/ml) Up to 10	0.300	<0.01**

Serum albumin, T.B Total bilirubin , ALP Alkaline phosphatase. D.B Direct bilirubin, AST Serum aspartate aminotransferase, AFP Alpha fetoprotein , ALT Serum alanine aminotransferase, PC % Prothrombin concentration GGT Serum gama glutamyle transferase, TP Total protein

Table (4): Pearson Correlation Matrix Between miRNA-155 And different variables in HCV NAIVE group

Studied variables	HCV NAIVE (N=20) MiR-155	
	r*	P-Value
AST(IU/L) Up to 40	0.218	>0.05
ALT(IU/L) Up to 42	0.132	>0.05
ALP (IU/L) M Up to 115 F Up to 104	0.425	>0.05
GGT (IU/L) 7 – 33	-0.014	>0.05
T.B (mg/dl) 0 – 1.0	0.447	>0.05
D.B (mg/dl) 0 – 0.25	-0.628	<0.05*
TP (g/dl) 6.5 – 8.5	-0.445	>0.05
Alb (g/dl) 3.5 - 5	0.237	>0.05
PC % 80 -120	0.227	>0.05
AFP (ng/ml) Up to 10	-0.30	>0.05

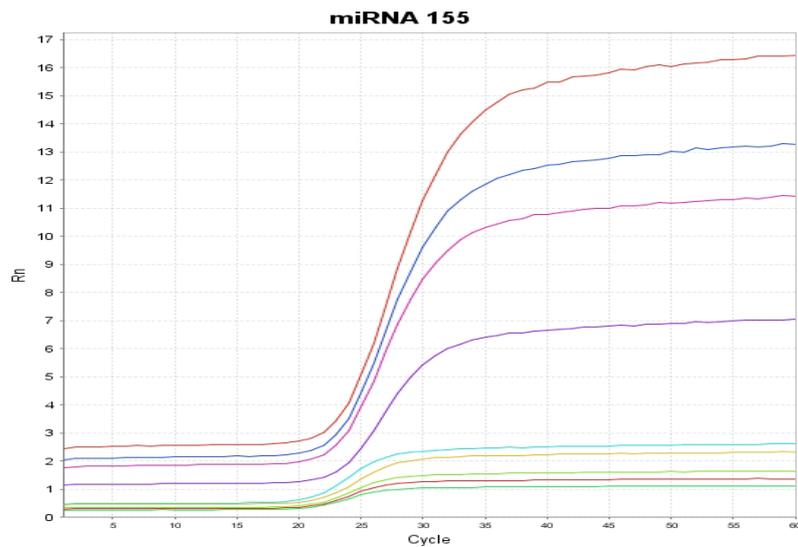


Figure 2: Amplification Plot (Rn vs. Cycle) of miRNA-155.

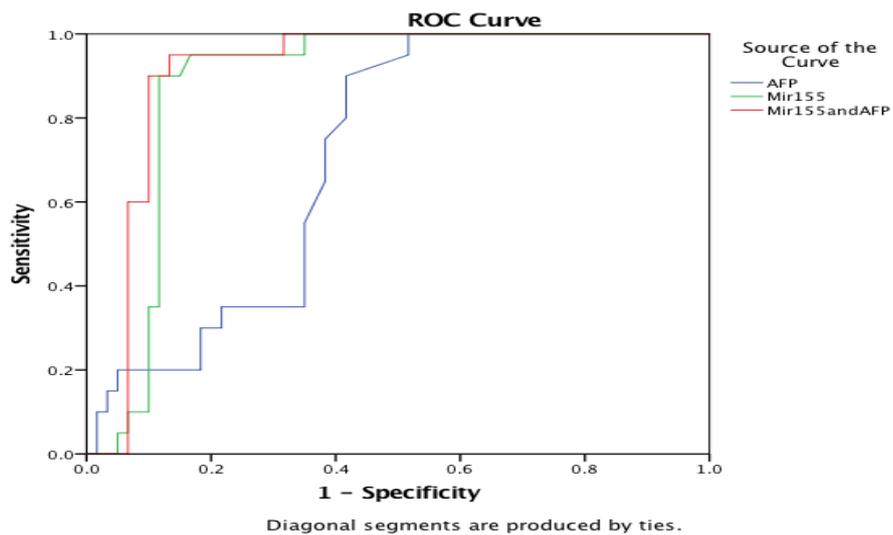


Figure 3: Receiver Operator of Characteristics (ROC) curve analysis of AFP and/or miRNA-155 in HCC group (No=20) versus non HCC .

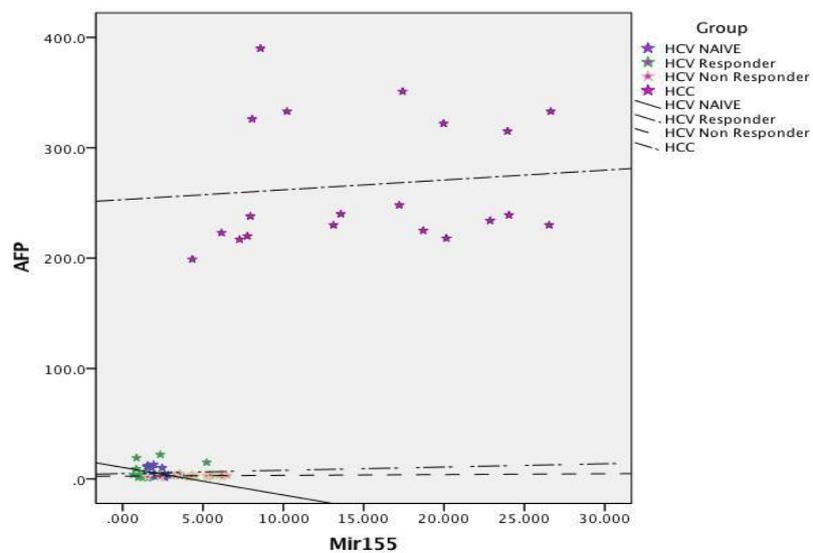


Figure 4: scatter plot with regression line showing correlation between miRNA-155 and AFP in HCC and HCV (NAIVE, responder and non responder) groups

Discussion

HCV is considered the most common etiology of HCC in Egypt. HCC is the common primary liver cancer with increasing incidence to become the 5th common malignancy worldwide and the third leading cause of cancer-related death. In Egypt, there was an increase in HCC incidence among chronic liver patients, HCC was reported to account for about 4.7% of chronic liver disease patients (15).

MicroRNAs (miRNAs) are approximately 22-nucleotide, noncoding, endogenous RNA molecules with an important role in a number of biological processes, including embryonic development, cell differentiation, and tumorigenesis(16). HCC as other malignancies is attributed to accumulated genetic alterations. As an oncomir, miRNA-155 gene was found to be over expressed in several solid tumors, such as thyroid carcinoma (17) breast cancer (18,19), cervical cancer (20) and lung cancer, where it is considered to be a marker of poor prognosis (21). To date the world literature has revealed that among the presently known miRNAs, miRNA-155 is one of the miRNAs most consistently involved in neoplastic diseases. Indeed, the frequently detected up-regulation of miRNA-155 in malignant cells allows to consider this gene predominantly as an oncogene playing a role in the pathogenesis of many human cancers, such as malignancies of the haematopoietic system (i.e. Hodgkin's Lymphoma, some types of Non Hodgkin's Lymphoma, AML, and CLL).

This work demonstrated that miRNA-155 is up regulated in HCC where mean RQ of circulating miRNA-155 in patients with HCC was significantly higher than patients with HCV

infected groups and healthy individuals, this was in agreement with Guan et al (22) and Hu et al (23).Who reported that miRNA-155 expression levels were enhanced in HCC tissues and this explained by

Circulating miRNAs originate from tumor tissues, and they are present in a certain form in the blood and are resistant to RNase activities (24). However, it remains unclear how circulating miRNAs originate from tumor tissues. It was suggested that miRNAs could be derived from dying or lysed tumor cells, invasive lymphoma cells, cells from tissues affected by long-term disease, or the active secretion of tumor cells (25).Other studies have investigated *miR-155* expression during hepatitis C virus (HCV) infection, and a positive correlation was found between the posttreatment persistence of HCV RNA in the serum and peripheral blood mononuclear cells (PBMCs) of infected patients and the expression of the *miR-155* precursor (BIC) in PBMCs (26). Another study has shown that HCV replication was positively correlated to the increased expression of mature *miR-155* in PBMCs of HCV-infected patients (27). Furthermore, *miRNA-155* was upregulated in liver tissues, serum and PBMCs of genotypes 1, 2 and 3 HCV-infected patients (28,29).

In the current study it was found that treatment-naïve patients with chronic HCV infection have increased expression of circulating miRNA-155. Importantly, miRNA-155 levels were decreased in patients who have successfully cleared HCV infection after therapy, suggesting a possible correlation between increased miRNA-155 and HCV viral presence and/or replication and this is agree with Bala et al (29).

Table(5): Sensitivity, Specificity, PPV, NPV and Accuracy of AFP and/or miRNA-155 in HCC group (No=20) versus non HCC (CLD and Control groups (No=80)).

The used marker	Sensitivity	Specificity	+ve predictive value	-ve predictive value	Accuracy
Plasma AFP at a cutoff point of (85.3) (ng/mL)	76.2 %	87.3%	90.2 %	72.41 %	81.0 %
MicroRNA-155 at cutoff point of (3.41) RQ	88.8 %	91%	92.4 %	89.5 %	91.4 %
Plasma AFP + MicroRNA-155	89.32 %	91.9 %	93.5 %	90.71 %	92 %

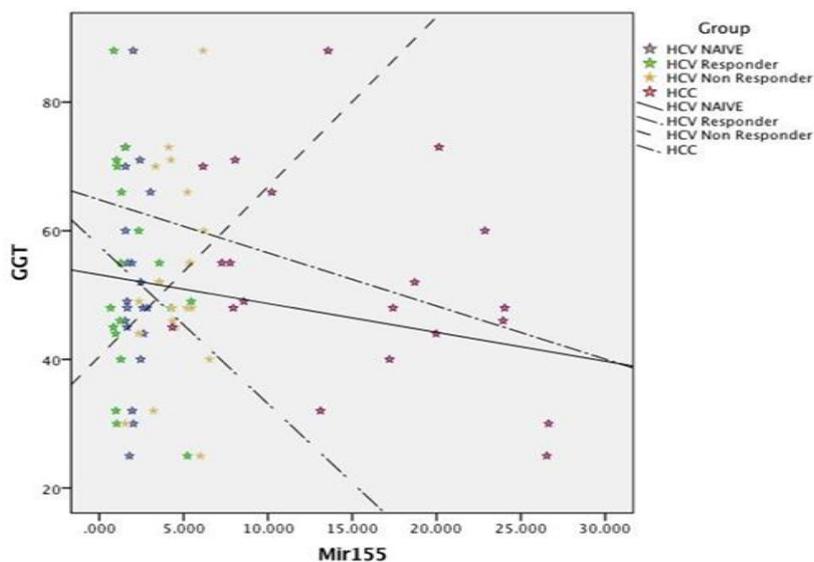


Figure 5:scatter plot with regression line showing correlation between miRNA-155 and GGT in HCC and HCV(NAÏVE,responder and non responder) groups

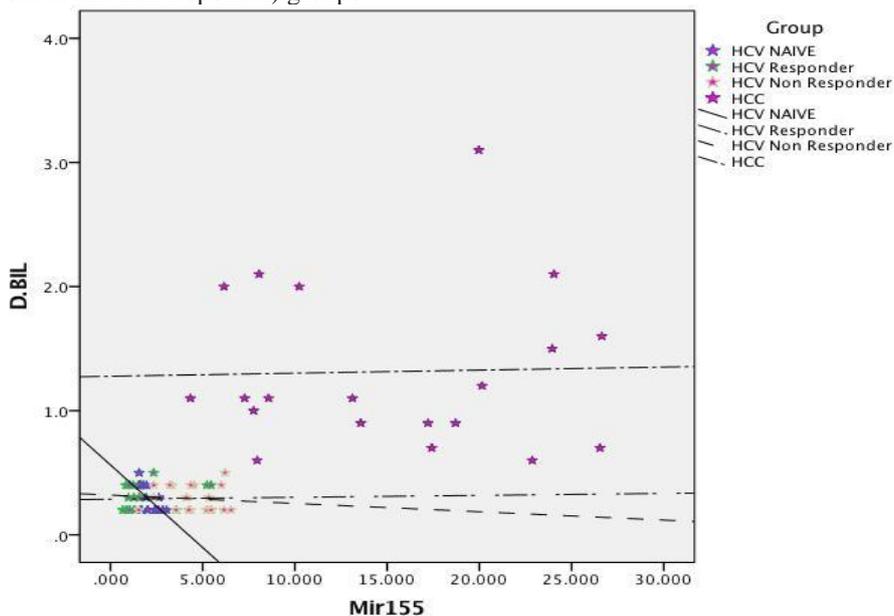


Figure 6:scatter plot with regression line showing correlation between miRNA-155 and D.Bil in HCC and HCV(NAÏVE, responder and non responder) groups

Recent studies suggest that miRNA-155 is not only limited to immune cells (dendritic cells, Kuffer cells, monocytes, NK cells, T cells), but also prevalent in non-immune cells (hepatocytes, endothelial cells) (29).

On the basis of these findings, it is most likely that both immune cells and hepatocytes contribute to increase of miRNA-155 in the circulation in HCV infection (29). Further studies are warranted to investigate the cellular source of circulating miRNAs.

In the current study, it is also observed that, the high expression of miRNA-155 was closely correlated with tumor size, TNM stage and vascular invasion, indicating that miRNA-155 may be involved in HCC progression, this was consistent with Hu et al (30) and song et al (31) who showed that miRNA-155 expression level were enhanced in HCC tissues also there was an association between miRNA-155 expression level, vascular invasion and tumor stage.

This is explained by many studies which have supported that high expression levels of miRNA-155 may be associated with high degree of malignancy and invasion in HCC. According to these data, miRNA-155 appears to be involved in tumor progression through the inhibition of multiple tumor suppressor genes, such as sex-determining region Y-gene related high mobility-group box gene (32,33) and suppressor in cytokine signaling1(34) thus promoting proliferation and invasion in HCC.

There is a close association between miRNA-155 expression levels and HCC prognosis (35,36). Huang et al., (37) demonstrated that there was an

association between miRNA-155 expression levels and 5-year relapse-free survival (RFS) of patients with HCC following radical surgical resection. The 5-year RFS of patients expressing high levels of miRNA-155 was reduced compared with patients exhibiting low levels of miRNA-155. It remains to be fully elucidated whether miRNA-155 is associated with early recurrence of HCC.

Regarding correlation between the mean relative quantity (RQ) of miRNA-155 and some studied parameters in this study, it was found that, a significant positive correlation between the mean RQs of miRNA-155 and AFP in HCC group was noticed.

In the current study There was significant correlation between miRNA-155 and each of GGT and AFP in HCC group. However, other reports demonstrated that no association was observed between miRNA-155 expression and α -fetoprotein levels of patients ($P > 0.05$) or the other studied variables such as the gender, age, tumor size, tumor number.

In the current study there was significant correlation between miRNA-155 and direct bilirubin in HCV naïve group. Also, there was significant correlation between miRNA-155 and each of AST and prothrombin concentration in HCV non responder group.

This study revealed that there was no significant correlation between ALT and miRNA-155 expression in HCC group and HCV patients groups and this in agree with Bala et al (29).

On the contrary de Bruijne et al., (38) demonstrated that miRNA-155 expression showed positive correlation with ALT and AST levels and

he suggested that this correlation may highlight the role of miRNA-155 in hepatic inflammation during GT4-cHCV infection.

Also this study revealed a significantly higher levels of AFP in HCC patients compared to the HCV naive patients, HCV responder patients, HCV non responder patients and control group. This was in agreement with Spadaro et al., (39) & Anwar et al., (40) who reported a significant elevation in serum AFP in HCC group compared to chronic liver patients and control group (39,40).

In the current study, the cutoff values and validity of serum AFP and circulating miRNA-155 for differentiation of HCC patients from those without HCC (CLD and controls) were determined by ROC curves.

From ROC curve analysis, the best cutoff of miRNA-155 chosen to differentiate HCC cases from non HCC subjects was 3.41RQs (fold change). and at this point the sensitivity, specificity, +ve predictive value(PPV), -ve predictive value(NPV), and Accuracy were 88.8%,91%, 92.4% ,89.5% ,91.4.% respectively and Thus, ROC curve analysis confirmed miRNA-155 as a valuable marker capable of discriminating the CLD patients, HCC and healthy individuals.

For AFP the best cutoff was 85.3ng/ml. at this cutoff point the Sensitivity, Specificity, PPV, NPV, Accuracy were (76.2% , 87.3 % , 90.2 % , 72.41 % , 81.0 %) respectively.

Furthermore, combined use of serum AFP and circulating miRNA-155for detection of HCC cases, had the advantage over the use of AFP alone as the sensitivity, specificity, PPV, NPP and

overall accuracy were increased (89.32 %, 91.9 %, 93.5%, 90.71 % and 92 % respectively).

Guan et al., (22)stated that compared with AFP, changes in circulating miRNA-155 are earlier and more accurately reflect the process of the formation of tumors than AFP. Also, circulating miRNA-155 was an independent significant factor for recurrence and was reported to be more sensitive than AFP for the detection of HCC.

The above results give an attention about the possibility of using miRNA-155 as a potential therapeutic target and this was previously stated by Guan et al (22) who suggested that miRNA-155 is related to the clinical characteristics of HCC and it may be a novel diagnostic marker and potential therapeutic target in HCC.

Conclusion:

miRNA-155 expression might be correlated with hepatitis C viral load and treatment response. Also, miRNA-155could be a novel diagnostic and prognostic biomarker for detection of HCC in combination with AFP as well as it could serve as a potential therapeutic target for HCV and HCC infection.

References

1. Choo QL, Kuo G, Weiner AJ, Overby LR, Bradley DW , Houghton M: Isolation of acDNA clone derived from a blood-borne non-A, non-B viral hepatitis genome. *Science* 244: 359-362, 1998.
2. Das S, Shetty RK, Kumar A, Shridharan RN, Tatineni R, Chi G, Mukherjee A ,Karande A: Monoclonal Antibodies against Hepatitis C Genotype 3aVirus Like Particle Inhibit Virus

- Entry in Cell CultureSystem. e53619:8(1),2013.
3. Li P, Lin Y, Zhang Y, Zhu Z ,Huo K: SSX2IP promote metastasis and chemotherapeutic resistance of hepatocellular carcinoma. *Journal of Translational Medicine* 11:52,2013.
 4. El-Zayadi AR, Badran HM, Shawky S, Emara S, El-Bareedy A , Sobhi M: Effect of surveillance for hepatocellular carcinoma on tumor staging and treatment decisions in Egyptian patients. *Hepato Int* 4:500-6,2010
 5. Mao B, Xiao H, Zhang Z, Wang D , Wang G: MicroRNA-21 regulates the expression of BTG2 in HepG2 liver cancer cells. *Molecular medicine reports* 12: 4917-4924,2015.
 6. Hu Q, Jiang H1, Su J1 ,Jiay Q: MicroRNAs as Biomarkers for Hepatocellular Carcinoma. A Diagnostic Meta-Analysis. *Clin. Lab* 59,2013.
 7. Faraoni I, Antonetti FR, Cardone J , Bonmassar E: "miR-155 gene a typical multifunctional microRNA". *BiochimBiophysActa* 1792:497–505,2009.
 8. TengG and Papavasiliou FN: _Silencing by microRNA-155. *Philosophical Transactions of the Royal Society B* 364 (1517): 631–7,2009.
 9. Mattiske S, Suetani RJ, NeilsenPM, Callen DF :The oncogenic role of miR-155 in breast cancer". *Cancer Epidemiol. Biomarkers Prev*21 (8): 1236–43,2012.
 10. Vargova K, Curik N, Burda P, Basova P, Kulvait V, Pospisil V, Savvulidi F, Kokavec J, Necas E, Berkova A, Obrtlikova P, Karban J, Mraz M, Pospisilova S, Mayer J, Trneny M, ZavadilJ,Stopka T:MYB transcriptionally regulates the miR-155 host gene in chronic lymphocytic leukemia..*Blood* 117 (14): 3816–25,2011.
 11. Corsten MF, Papageorgiou A, Verhesen W, Carai P, Lindow M, Obad S, Summer G, Coort SL, Hazebroek M, van Leeuwen R, Gijbels MJ, Wijnands E, Biessen EA, De Winther MP, Stassen FR, Carmeliet P, Kauppinen S, Schroen B, Heymans S. :MicroRNA profiling identifies microRNA-155 as an adverse mediator of cardiac injury and dysfunction during acute viral myocarditis. *Circ; Res.*111 (4): 415–25,2012.
 12. Zhang C, Huys A, Thibault PA,Wilson JA: Requirements for human Dicer and TRBP in microRNA-122 regulation of HCV translation and RNA abundance. *Virology* 433:479–488,2012.
 13. Schuppan D, Krebs A, Bauer M , Hahn EG: Hepatitis C and liver fibrosis. *Cell Death Differ.* 10 Suppl 1:S59–S67,2003.
 14. Dolganiuc A, Norkina O, Kodys K, et al: Viral and host factors induce macrophage activation and loss of toll-like receptor tolerance in chronic HCV infection. *Gastroenterology* 133(5):1627–1636,2007.
 15. Ezzat, H., Lotfy, A., Alalfy, M., El-Taher, S., Mokhtar, A., Mohamed, S., , EL-Senousy, F:The Significance of Circulating Micro RNA-122 as a Non Invasive Diagnostic Marker of Liver Injury in Egyptian Chronic Hepatitis C virus Infected and Cirrhotic Patients with and without Hepatocellular Carcinoma. *Clinical Medicine and Diagnostics* 4(1): 1-8,2014.
 16. Hu Q, Jiang H, Su J ,Jiay Q: MicroRNAs as Biomarkers for Hepatocellular Carcinoma. A Diagnostic Meta-Analysis.*Clin. Lab* 59,2013.
 17. Nikiforova,M.N,Tseng,G.C,Steward,D.,Diorio ,D.,,Nikiforov,Y.E:Micro-RNA expression profiling of thyroid tumors.*biological*

- significance and diagnostic utility. *J Clin Endocrinol Metab* 93:1600-1608,2008.
18. Volinia, S., Calin, G.A., Liu, C.G., Ambs, S., Croce C.M: A micro RNA expression signature of human solid tumors defines cancer gene targets. *Proc Natl Acad Sci USA* 103:2257-2261,2006.
 19. Iorio MV, Ferracin M, Liu CG, Veronese A, Spizzo R: MicroRNA gene expression deregulation in human breast cancer. *Cancer Res* 65:7065-7070,2005.
 20. Wong TS, Liu XB, Wong BY, Ng RW, Yuen AP, et al: Mature miR-184 as a potential oncogenic microRNA of squamous cell carcinoma of tongue. *Clin Cancer Res* 14:2588-2592,2008.
 21. Yanaihara N, Caplen N, Bowman E, Seike M, Kumamoto K, Yi M, Stephens R, Okamoto A, Yokota J, Harris C: Unique microRNA molecular profiles in lung cancer diagnosis and prognosis. *Cancer Cell* 9:189-198,2006.
 22. Guan C, Yang F, He X, Li T, Yang Q, He H, Xu M: miR-155 expression and clinical significance in HCC. *Oncol Lett* 11(2): 1574-1580,2016.
 23. Hu YH, Cai Q, Lan QY: Expression of miR-155 in hepatocellular carcinoma and its effect on the proliferation of hepatocellular carcinoma cells. *World J Gastroenterol* 19:1737-1741,2012.
 24. Mitchell PS, Parkin RK, Kroh EM, Fritz BR, Wyman SK, Pogosova-Agadjanyan EL, Peterson A, Noteboom J, O'Brian KC, Allen A, Lin DW, Urban N, Drescher CW, Knudsen BS, Stirewalt DL, Gentleman R, Vessella RL, Nelson PS, Martin DB, Tewari M: Circulating microRNAs as stable blood-based markers for cancer detection. *Proc Natl Acad Sci U S A* 105: 10513-10518,2008.
 25. Wang X, Zhang J, Zhou L, Lu P, Zheng ZG, Sun W, Wang JL, Yang XS, Li XL, Xia N, Zhang N, Dou KF: Significance of serum microRNA-21 in diagnosis of hepatocellular carcinoma (HCC): clinical analyses of patients and an HCC rat model. *Int J Clin Exp Pathol* 8(2):1466-1478,2015.
 26. Sidorkiewicz M, Grek M, Jozwiak B, Majda-Stanislawski E, Piekarska A, Bartkowiak J: Expression of microRNA-155 precursor in peripheral blood mononuclear cells from Hepatitis C patients after antiviral treatment. *Acta Virol* 54:75-78,2010.
 27. Grek M, Piekarska A, Bartkowiak J, et al: Coordinated increase of miRNA-155 and miRNA-196b expression correlates with the detection of the antigenomic strand of hepatitis C virus in peripheral blood mononuclear cells. *Int J Mol Med* 28:875-880,2011.
 28. Zhang Y, Wei W, Cheng N, Wang K, Li B, Jiang X, Sun S: Hepatitis C virus-induced up-regulation of microRNA-155 promotes hepatocarcinogenesis by activating Wnt signaling. *Hepatology* 56:1631-1640,2012.
 29. Bala S, Tilahun Y, Taha O, Alao H, Kodys K, Catalano D, Szabo G: Increased microRNA-155 expression in the serum and peripheral monocytes in chronic HCV infection. *J Transl Med* 10:151,2012.
 30. Hu YH, Cai Q, Lan QY: Expression of miR-155 in hepatocellular carcinoma and its effect on the proliferation of hepatocellular carcinoma cells. *World J Gastroenterol* 19:1737-1741,2012.

31. Song CG, Wu XY, Fu FM, et al: Correlation of miR-155 on formalin-fixed paraffin embedded tissues with invasiveness and prognosis of breast cancer. *Zhonghua Wai Ke Za Zhi* 50:1011–1014,2012.
32. Michieli P, Chedid M, Lin D, Pierce JH, Mercer WE, Givol D. Induction of WAF/CIP1 by Ap53- independent pathway. *Cancer Res* .1994;54:3391-3395.
33. Xie Q, Chen X, Lu F, et al: Aberrant expression of microRNA 155 may accelerate cell proliferation by targeting sex-determining region Y box 6 in hepatocellular carcinoma. *Cancer* 118:2431-2442,2012.
34. Yan XL, Jia YL, Chen L, et al: Hepatocellular carcinoma –associated mesenchymal stem cells promote hepatocarcinoma progression. Role of the S100A4-miR-155-SOCS1-MMP9 axis. *Hepatology* 57:2274-2286,2013.
35. Braconi C, Henry JC, Kogure T, Schmittgen T, Patel T: The role of microRNAs in human liver cancers. *Semin Oncol* 38:752-763,2011.
36. Shrivastava S, Steele R, Ray R, Ray RB. microRNA : Role in hepatitis C virus pathogenesis. *Genes Dis* 2:35-45,2015.
37. Huang YH, Lin KH, Chen HC, et al: Identification of postoperative prognostic microRNA predictors in hepatocellular carcinoma. *journal.pone* 188:10-.1371,2012.
38. de Bruijne J, Buster EH, Gelderblom HC, et al. Treatment of chronic hepatitis C virus infection. Dutch national guidelines 66:311–322,2008.
39. Spadaro A, Ajello A, Morace C, Zirilli A, D'Arrigo G, Luigiano C, Martino F, Bene A, Migliorato D, Turiano S : Serum chromogranin-A in hepatocellular carcinoma diagnostic utility and limits. *World J Gastroenterol* 11:1987-90,2005.
40. Anwar WA, Khaled HM, Amra HA, El-Nezami H, Loffredo CA : Changing pattern of hepatocellular carcinoma (HCC) and its risk factors in Egypt. possibilities for prevention. *Mutat Res* 659:176-84,2008.